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2 following a high-intensity interval training session.

3

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28

29 **Abstract**

30 **Purpose:** To assess the concurrent validity of a continuous blood glucose monitoring system
31 (CGM) Post-Breakfast, Pre-exercise, Exercise and Post-exercise, while assessing the impact of
32 two different breakfasts on the observed level of validity. **Methods:** Eight non-diabetic
33 recreational athletes (age: 30.8±9.5 years; height: 173.6±6.6 cm; body mass: 70.3±8.1 kg) took
34 part in the study. Blood glucose concentration was monitored every 10 min using both a CGM
35 (FreeStyle Libre, Abbott, France) and finger-prick blood glucose measurements (FreeStyle
36 Optimum, Abbott, France) over 4 different periods (Post-Breakfast, Pre-Exercise, Exercise and
37 Post-Exercise). Two different breakfasts (carbohydrates- [CHO] and protein- [PROT] oriented)
38 over two days (2x2 days in total) were used. Statistical analyses included the Bland-Altman
39 method, standardized mean bias (expressed in standardized unit), median absolute relative
40 difference (MARD) and the Clarke Error Grid (EGA). **Results:** Overall, mean bias was trivial-
41 to-small at Post-Breakfast (effect size ± 90% confidence limits: -0.12±0.08), Pre-Exercise (-
42 0.08±0.08) and Post-Exercise (0.25±0.14), while moderate during Exercise (0.66±0.09). Higher
43 MARD was observed during Exercise (13.6% vs 7 to 9.5% for the other conditions). While
44 there was no effect of the breakfast type on the MARD results, EGA revealed higher value in
45 Zone D (*i.e.* clinically unsafe zone) during Exercise for CHO (10.5%) compared with PROT
46 (1.6%). **Conclusion:** The CGM device examined in this study can only be validly used at rest,
47 after both a CHO and PROT-rich breakfast. Using CGM to monitor blood glucose concentration
48 during exercise is not recommended. Moreover, the accuracy decreased when carbohydrates
49 are consumed before exercise.

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59 **Introduction**

60 Regulation of blood glucose has first been widely studied from a health perspective.
61 Hyperglycemia for example, is believed to be an independent risk factor for the development
62 of several diseases such as type II diabetes mellitus¹ and cardiovascular disease.² More recently,
63 the monitoring of blood glucose concentration has also elicited great interest in sport, as
64 hypoglycaemia influences both physical and cognitive performances.³

65 In particular, it is known that at the beginning of exercise or after half-time in team sports,
66 athletes experience transient hypoglycemia, which may affect physical and cognitive
67 performance.⁴ Moreover, it has then been shown that a large glycemic variability exists among
68 individuals in the general population.⁵ Additionally, similar results have been shown in sub-
69 elite athletes,⁶ suggesting that providing more individualized guidelines to regulate blood
70 glucose would be beneficial for both health and performance goals.

71 The emergence of new technologies such as continuous glucose monitoring (CGM) devices has
72 allowed blood glucose concentration dynamics to be captured more frequently and less
73 invasively than traditional measures such as finger pricks. Indeed, as CGM devices only need
74 to be placed once (usually on the back of the arm), it can be used for several days without
75 disturbing sport practices. So far, these devices have been mainly used by diabetic populations
76 but as the technology becomes more accurate, less invasive, and less expensive, their use has
77 increased in other populations and especially in healthy individuals. Therefore, the inclusion of
78 CGM in sport nutritionists' monitoring tool box could help to optimize nutritional strategies
79 before and during exercise, and in turn, improve athletes' performance by preventing
80 hypoglycemia. However, to date, the validity of these new systems at rest or during exercise
81 has been only assessed in diabetics patients and showed promising results.⁷ Evidence regarding
82 its relevance with an athletic population is still lacking. Moreover, the ability of such devices
83 to detect potential glucose fluctuations due to different nutritional intakes need to be confirmed.

84 Therefore, the aim of this study was to assess the concurrent validity of a new CGM device
85 during different periods, *i.e.* pre, during and after exercise, while assessing the potential impact
86 of different nutritional intakes in the observed level of validity.

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90 **Methodology**

91 **Study Population**

92 Eight non-diabetic recreational athletes (5 females, 3 males) (age: 30.8 ± 9.5 years; height:
93 173.6 ± 6.6 cm; body mass: 70.3 ± 8.1 kg) who regularly participate in running and resistance-
94 based training (8 ± 2 hours per week) were included in the study. An a priori power analysis was
95 conducted using the package *pwr* from R software (Version 4.0.0) for t-tests for non-parametric
96 data with a significance level alpha of 0.05 a power of 0.8 and add a non-parametric correction
97 of 15%. Result showed a minimal sample of 310 paired observations for 8 participants were
98 necessary. Alcohol intake was prohibited during the study period. Regarding female
99 participants, we ensured they were all within the same menstrual phase during the study period.

100 Participants provided informed consent prior to starting the study. Ethics approval was granted
101 before any data collection was undertaken and the recommendations of the Declaration of
102 Helsinki were respected.

103

104

105 **Design**

106 A concurrent validity design was employed to assess the validity of a CGM system against
107 finger prick measures which was considered as the reference method. Over 2 consecutive
108 weeks, participants took part in 4 nonconsecutive standardized days. Each standardized day was
109 broken-down into 4 distinct periods: 1) Post-Breakfast which corresponded to the first hour
110 after the end of the Breakfast 2) Pre-Exercise which corresponded to the first hour following
111 the Post-Breakfast, 3) Exercise, which started 2 hours after the end of the breakfast and lasted
112 from the beginning of the warm up to the end of the workout and 4) Post-exercise, which started
113 immediately at the end of the workout, and up to 30 min later. A detailed outline of the
114 standardized day structure is provided in Figure 1. Nutritional intake during breakfast was
115 manipulated in order to provide either a high carbohydrate (CHO) or protein (PROT) breakfast,
116 to induce different levels of resting and pre-exercise glycemia. Each typical breakfast was
117 repeated twice. Over those standardized days, blood glucose was measured continuously with
118 a CGM, while finger prick measures were taken every 10 minutes and. Day 1 was used for each
119 participant to familiarize with the CGM and ensure calibration (as per manufacturer
120 recommendations) before the experimentation could start. Between day 2 and 13, participants

121 undertook at their convenience the 4 standardized days. They were also instructed to have at
122 least one full day of recovery between each experimental day.

123

124 **Insert Figure 1**

125

126 **Methodology**

127 *Continuous glucose monitoring.* Each participant was provided with a CGM system (FreeStyle
128 Libre, Abbott, France) over the full duration of the study. Each participant inserted a sensor
129 (FreeStyle Libre, Abbott, France) in their non-dominant upper arm (*i.e.* back the triceps
130 brachialis) one day before the beginning of the study. Glucose concentration was recorded in
131 the interstitial fluid every minute.

132 *Finger prick blood glucose.* Finger prick (FreeStyle Optium, Abbott, France) measures were
133 collected following the procedure described by Gomez.⁸ Each sample was immediately
134 analysed using the FreeStyle Libre reader (FreeStyle Libre Reader, Abbott, France) (The
135 validity and reliability of this device has been previously confirmed.⁹

136 *Breakfast.* Two typical breakfasts were employed. The CHO breakfast contained a high
137 proportion of carbohydrates (CHO) with 1 g·Kg⁻¹ of body mass with a ceiling set at 70g of
138 carbohydrates per breakfast (*e.g.* breakfast contained a mix of orange juice, bread and jam). The
139 macronutrients and energy were as follow: 65±7g of carbohydrates, 9±1g of proteins and 1±0g
140 of fat for a total of 311±31 Kcal. The protein (PROT) breakfast was isoenergetic compared with
141 CHO (*e.g.* breakfast contained a mix of eggs, ham and cheese). The macronutrients and energy
142 were as follow: 1±0g of carbohydrates, 30±0g of proteins and 23±0g of fat for a total of 311±31
143 Kcal.

144 *Standardized exercise.* Participants completed the 30-15 Intermittent Fitness Test (30-15_{IFT}) as
145 described by Buchheit et al.¹⁰ prior the beginning of the study. The speed (km·hr⁻¹) achieved
146 by each participant during the last successfully completed stage of the test was recorded (V_{IFT})
147 in order to prescribe exercise intensity. The standardized exercise started with a 10-min low-
148 intensity run (30 to 40% of V_{IFT}) and was followed by a high-intensity intermittent training
149 exercise performed outdoor. The trials consisted of six reps of 3-min running intervals
150 interspersed with 2 min of passive recovery. Reps 1 and 2 were performed at 75% V_{IFT}, reps 3
151 and 4 at 80% V_{IFT} and reps 5 and 6 at 85% V_{IFT}. The session was ended with a 10-min walk.

152 *Data processing.* Each time point within a specific period was averaged as described above to
153 perform the concurrent validity analysis for each method (CGM and finger prick) and per
154 specific period (Figure 1). Each standardized day was analyzed first without (overall) and then
155 as a function of breakfast type (CHO and PROT).

156 **Statistical Analysis**

157 Bland-Altman method for repeated measures and standardized mean bias were first applied to
158 assess the agreement between CGM and finger prick measures at each specific period.¹¹ The
159 following thresholds were applied to rate the magnitude of the bias as follow: >0.2 (small), >0.6
160 (moderate), >1.2 (large) and >2 (very large).¹²

161 Additionally, analysis of the median average relative difference (MARD)¹³ and the Clarke Error
162 Grid Analysis (EGA)¹⁴ were conducted. Regarding MARD, further comparisons between the
163 different periods were performed using Wilcoxon test and/or Kruskal-Wallis tests. Level of
164 statistical significance was set at $P < 0.05$. Results were further analyzed while calculating
165 standardized differences, *i.e.* Wilcoxon effect sizes. The thresholds to rate the magnitude of
166 the effects were the same than those used for mean bias. Regarding EGA, results were divided
167 into 5 zones (A, B, C, D, E). Each zone denotes a degree of clinical implications of blood
168 glucose concentration measures. Zones A and B were considered clinically acceptable while
169 zone C, D and E (erroneous treatment) were deemed possibly unsafe.¹⁴

170

171 **Results**

172 The Bland-Altman analysis for the 4 periods is presented in Figure 2 and reported as mean bias
173 (standard error). Irrespectively of the breakfast content, mean biases were trivial-to-small for
174 Post-Breakfast (-2.99 [17.75] mg/dL), Pre-Exercise (-1.67 [10.95] mg/dL), Post-Exercise (4.18
175 [17.88] mg/dL) and moderate during Exercise (12.25 [13.86] mg/dL). Regarding CHO
176 breakfast, mean biases were trivial-to-small for Post-Breakfast (-1.43 [25.98] mg/dL), Pre-
177 Exercise (-4.29 [11.66] mg/dL), Post-Exercise (3.32 [18.18] mg/dL) and moderate during
178 Exercise (14.06 [13.81] mg/dL). For PROT Breakfast, trivial mean bias was observed for Pre-
179 Exercise (0.91 [8.98] mg/dL), Post-Breakfast (-4.51 [8.31] mg/dL) and Post-Exercise (5.13
180 [15.98] mg/dL), while moderate mean biases were observed for Exercise (10.47 [13.19]
181 mg/dL).

182

183 **Insert Figure 2**

184 **Insert Figure 3**

185 The results of the MARD analysis between the different periods are presented in Table 1 and
186 2.

187
188 **Insert Table 1 and 2**

189
190 Results regarding EGA are presented in Table 3. Irrespectively of the breakfast content, Post-
191 Breakfast, Pre-Exercise, and Post-Exercise periods fell into Zone A (accurate) and B (benign
192 errors) (100%). However, during Exercise, 94% of the values fell into A (70.4%) and B
193 (23.6%), and 6% in Zone D (failure to treat errors). For CHO breakfast, 10.5% of data fell into
194 Zone D for Exercise, while the other periods fell into Zone A and B. Similarly, for PROT
195 breakfast, 1.6% fell into Zone D during the Exercise period.

196
197 **Insert Table 3**

198 **Discussion**

199 The aim of this study was 1) to investigate the concurrent validity of a new CGM device in
200 recreational athletes at Post-Breakfast, Pre-exercise, Exercise and Post-exercise, and 2) to
201 assess the potential impact of either a CHO-rich or protein-rich breakfast on the observed level
202 of validity. The main results highlighted that, while the validity of CGM was acceptable at rest
203 (*i.e.* Post-Breakfast, Pre-Exercise and Post-Exercise), it was lower during Exercise and
204 especially after the CHO breakfast.

205 The first results demonstrated trivial-to-small mean bias during all the non-exercise periods,
206 irrespectively of nutritional intake. Moreover, all results from EGA fell into the “clinically safe
207 zone” (A and B), albeit during Exercise. These results are similar to those shown previously in
208 non-athletic diabetic populations.¹⁵ Indeed, the present results suggest that assessing glucose
209 dynamics at rest is feasible with this CGM device. This could open the door to a better
210 individualization of nutritional strategies.⁵

211 Yet, we observed a higher bias during Exercise compared with the other periods, confirming
212 previous studies in a non-athletic diabetic population.¹⁶ Reasons that may contribute to the
213 reduced validity of the CGM device in this context include microcirculation perturbations as a
214 as a result of movements around or within the insertion area, increases in body temperature and
215 rapid fluxes in glucose levels during exercise.¹⁷ Regarding the likely physiological time lag of
216 glucose transport between blood and interstitial fluid compartments (see Figure 3, finger pricks
217 measures changed faster Post-Breakfast than that of the CGM device), it should be noted that
218 it might not have accounted for the observed difference in accuracy as the pattern is not only
219 delayed but it varies with time and conditions. Indeed, while a clear hypoglycemia was observed
220 with finger prick measures immediately at the start of exercise (which was the expected
221 physiological response), the CGM showed an increased blood glucose response (Figure 2).
222 Nonetheless, this discrepancy indicates that the CGM device was unable to detect a potential
223 hypoglycemia observed at the onset of exercise, and could therefore not be used to assess
224 strategies aiming at preventing this phenomenon in practice. It is worth mentioning that a trend
225 for a better agreement was observed toward the end of the exercise periods (Figure 2). If the
226 duration of the exercise also affects the accuracy of CGM, it means that while the device may
227 not be suitable for sport including short and intermittent exercise durations, its use could
228 perhaps be considered during longer event such as cycling, trail or triathlon. This potential
229 better accuracy toward longer exercise duration highlights the need to conduct further research
230 involving 1) longer exercise duration, 2) nutritional intake during long endurance race 3)
231 various exercise modalities and 4) different intensities.

232 To examine the potential effect of the absolute levels of glycemia on the validity of the CGM
233 device, different breakfasts were proposed (CHO and PRO). Similar MARD and EGA results
234 were observed, suggesting that the CGM validity was not affected by the breakfast content
235 during non-exercise periods (*i.e.* Post-Breakfast, Pre-Exercise, Post-Exercise). Specific pre-
236 competition nutritional strategies can have a positive influence on both the acute running
237 performance among rugby league players¹⁸ or endurance athletes,¹⁹ and the chronic training
238 adaptations to training.²⁰ Consequently, the use of this CGM device could be considered by
239 practitioners willing to monitor glycemic responses before and after competition or training, to
240 ensure the efficacy of the nutritional strategies employed.

241 However, during the Exercise period, the CGM accuracy was modulated by breakfast content.
242 Indeed, a 10 times higher value in Zone D of the EGA (*i.e.* clinically unsafe) was observed post
243 CHO (10.5%) compared with post PROT (1.6%) breakfast. In our study, zone D corresponds

244 to the situation where finger prick measures indicate an hypoglycemic state whereas CGM
245 measures are within the normal range¹⁴ suggesting that CGM failed to detect the hypoglycemia
246 occurring during exercise after the CHO-rich breakfast. It is well known there is a rapid drop
247 of blood glucose concentration at the onset of exercise, due to an increased glucose uptake by
248 exercising muscles.²¹ This physiological mechanism could explain why the sensor lacks
249 sensitivity to rapid changes in glucose concentration, as observed in the present study. As it
250 stands, if practitioners want to monitor blood glucose during high-intensity intermittent
251 exercise, they need to consider other devices than CGM (e.g. finger prick).

252

253 **Practical applications**

254

255 - The present CGM system provided valid measures at rest. Therefore, the use of such a
256 system may allow for a better individualization of nutritional strategies before or after
257 competition.

258

259 - The level of validity was lower during high-intensity intermittent training and was in
260 addition influenced by the type of breakfast consumed (i.e. high carbohydrates or high
261 protein). Consequently, practitioners should avoid using this device during intermittent
262 exercise.

263

264 **Conclusion**

265 Daily monitoring of blood glucose is of importance in athletes given the likely impact of
266 glycemia on performance and the individualized nutritional recommendations that can be made
267 with CGM. Our results highlighted that the CGM device examined in the present study
268 presented only trivial-to-small bias when compared with a traditional fingerpick device at rest,
269 suggesting that it could be used confidently during this specific period. The CGM device is not
270 valid enough to monitor glucose during intermittent exercise. Further analyses should however
271 evaluate the validity of this device over longer exercise duration.

272

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338

339

Table and figure caption

Figure 1. Schematic representation of the study design.

Figure 2. Bland-Altman analysis between the continuous glucose monitoring device (CGM) and finger prick measures (FPBG). Dash lines represent the limits of agreements.

Figure 3. Continuous glucose monitoring (CGM) and finger prick measures during each standardized condition, when ingesting a carbohydrate- (upper) and protein- (lower) oriented breakfasts, with the 2 days of each breakfast condition pooled for each participant ($n = 2 \times 8$ for each curve). Data are presented as mean (SE).

Table 1. Median Absolute Relative Difference between the continuous glucose monitoring device (CGM) and finger prick measures. Data are median (interquartile range) and expressed in percentage. *: significantly different from Post-Breakfast. #: significantly different from Pre-Exercise. †: significantly different from Exercise. Comparisons between period are presented as effect size with 90% confidence interval.

Table 2. Comparisons between period are presented as effect size for Wilcoxon test with 90% confidence interval.

Table 3. Clark Error Grid Analysis between the continuous glucose monitoring device (CGM) and finger prick measures. Zone A represents a clinically accurate measure. Zone B stands for benign errors. Zone C represents overcorrection errors. Zone D and E represent failure to treat errors and erroneous treatment errors respectively. For more details see Clarke et al. (1987).

Table 1

	Post-Breakfast	Pre-Exercise	Exercise	Post-Exercise
Overall	9.1 (4.6-13.8)	7.1 (3.6-13.4) #	13.6 (6.8-23.2)*	9.4 (5.0-17.3) #†
CHO	9.4 (5.3-16.8)	7.1 (3.9-13.2) *	16.2 (7.4-25.6) *#	10.1 (6.1-16.9) #†
PROT	8.8 (4-11.9)	7.0 (3.4-13.4)	11.3 (6-19.7) *#	8.2 (4.1-17.3)

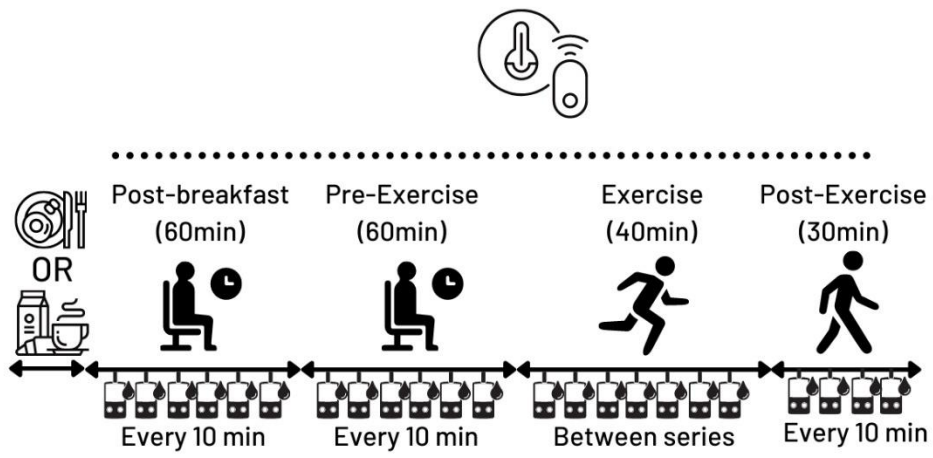
Table 2

	Post-Breakfast vs. Exercise	Post-Breakfast vs. Post-Exercise	Pre-Exercise vs. Exercise	Pre-Exercise vs. Post-Exercise	Exercise vs. Post Exercise
Overall	0.24 (0.17 to 0.31)	0.07 (0.01 to 0.16)	0.31 (0.24 to 0.38)	0.16 (0.07 to 0.24)	0.15 (0.06 to 0.23)
CHO	0.24 (0.13 to 0.34)	0.06 (0.01 to 0.18)	0.37 (0.27 to 0.46)	0.19 (0.07 to 0.31)	0.18 (0.07 to 0.28)
PROT	0.24 (0.14 to 0.34)	0.08 (0.01 to 0.2)	0.26 (0.16 to 0.36)	0.18 (0.01 to 0.24)	0.12 (0.02 to 0.24)

Table 3

	Zone	Post-Breakfast	Pre-Exercise	Exercise	Post-Exercise
Overall	A (Accurate)	189 (88.3%)	213 (93.4%)	176 (70.4%)	100 (76.3%)
	B (Benign errors)	25 (11.7%)	14 (6.1%)	59 (23.6%)	31 (23.7%)
	D (Failure to treat errors)	/	1 (0.5%)	15 (6.0%)	/
CHO	A (Accurate)	85 (80.2%)	104 (92.0%)	81 (65.3%)	52 (75.4%)
	B (Benign errors)	21 (19.8%)	9 (8.0%)	30 (24.2%)	17 (24.7%)
	D (Failure to treat errors)	/	/	13 (10.5%)	/
PROT	A (Accurate)	104 (96.3%)	109 (94.8%)	95 (75.4%)	48 (77.4%)
	B (Benign errors)	4 (3.7%)	5 (4.3%)	29 (23.0%)	14 (22.6%)
	D (Failure to treat errors)	/	1 (0.9%)	2 (1.6%)	/

Figure 1



Legend:



Protein Breakfast



Carbohydrate Breakfast



Continuous glucose monitoring



Finger prick blood glucose

Figure 2

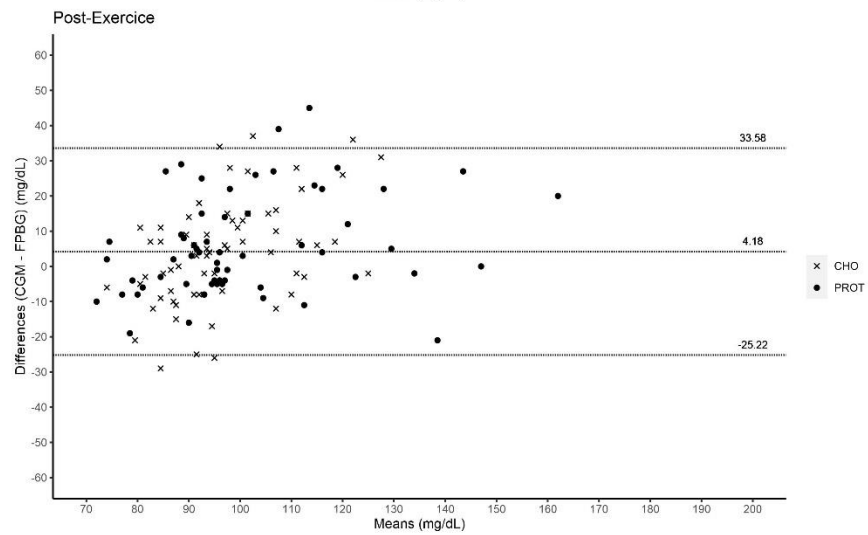
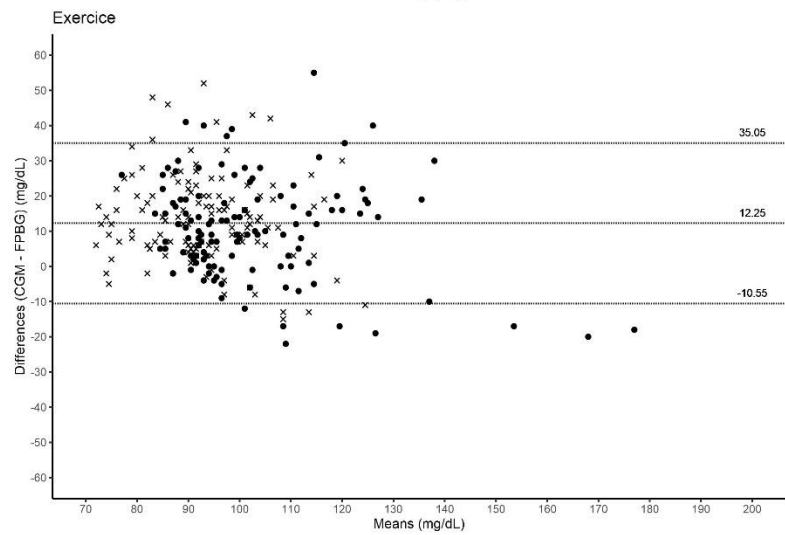
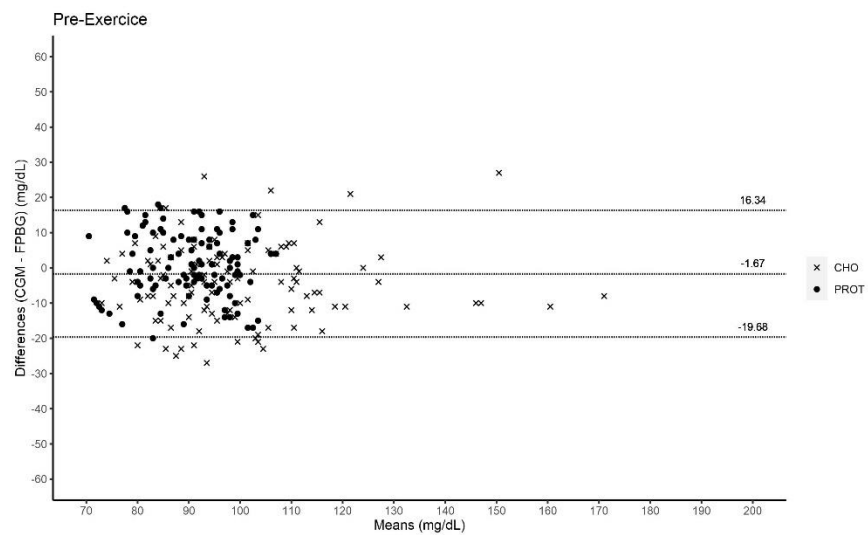
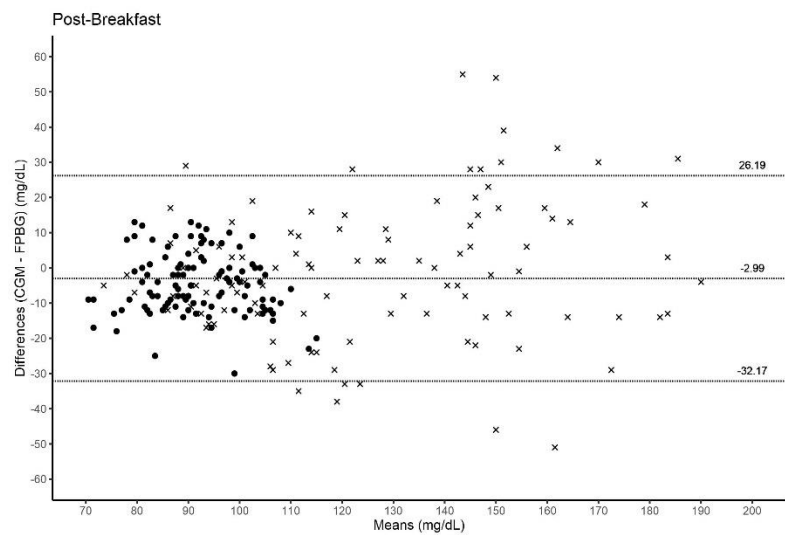


Figure 3

